

Marine Environmental Appraisal of an Ocean Energy Test Site in Inner Galway Bay

Produced by

AQUAFACT International Services Ltd

On behalf of

Hydraulics & Maritime Research Centre April 2010

AQUAFACT INTERNATIONAL SERVICES ltd 12 KILKERRIN park TUAM rd GALWAY city www.aquafact.ie

info@aquafact.ie

tel +353 (0) 91 756812 fax +353 (0) 91 756888

Table of Contents

| 2. Marine Benthos | |
|---|----|
| | 4 |
| 2.1. Sampling Procedure & Processing | |
| 2.2. Data Processing | 6 |
| 2.3. Results | 9 |
| 2.3.1. Fauna | |
| 2.3.1.1. UNIVARIATE ANALYSES | 9 |
| 2.3.1.2. MULTIVARIATE ANALYSES | 10 |
| 2.3.2. Sediment | 13 |
| 2.3.3. Underwater imaging | 17 |
| 2.3.3.1. Sediment Profile Imaging (SPI) | 17 |
| 2.3.4. Discussion | 17 |
| 3. Marine reptiles, mammals and birds | 19 |
| 3.1. Marine reptiles of Ireland's Atlantic margin | 19 |
| 3.2. Marine Mammals of Ireland's Atlantic Margin | 20 |
| 3.3. Birds of Ireland's Atlantic Margin | 22 |
| 3.3.1. Inner Galway Bay | |
| 4. References | 28 |

List of Figures

| Figure 1: Location of the ocean energy site within Galway Bay2 |
|--|
| Figure 2: Ocean Energy Buoy moored in Galway Docks3 |
| Figure 3: Ocean Energy Buoy moored in Galway Bay3 |
| Figure 4: Map showing sampling locations4 |
| Figure 5: Dendrogram showing each station from the 6 stations sampled in the ocean energy test |
| site |
| Figure 6: MDS ordination showing each station from the 6 stations sampled in the ocean energy test |
| site |
| Figure 7: Sediment grain size data15 |
| Figure 8: Sediment type according to Folk (1954)16 |
| Figure 9: Map showing Wave energy test site and areas where cetaceans were recorded between |
| 2008 and 2010 |

List of Tables

| able 1: Station co-ordinates | 5 |
|---|-----|
| able 2: The classification of sediment particle size ranges into size classes (adapted from Buchana | an, |
| 1984) | 6 |
| able 3: Diversity indices for the 6 stations sampled at the ocean energy test site. | 9 |
| able 4: Granulometry results for the 6 stations sampled at ocean energy test site in Galway Bay. | 14 |
| able 5: Number of Species, Individuals, their Richness, Evenness and Diversity at each Station | 18 |
| able 6: Mean number of water birds in the area from 1996 to 2000 | 23 |
| able 7: Concentrations of waterbirds in Inner Galway Bay between 1994/95 and 2000/01 | 25 |

List of Appendices

| Appendix 1 | Faunal Abundance |
|------------|-------------------------|
| Appendix 2 | Underwater photographs |
| Appendix 3 | Sediment Profile Images |

1. Introduction

AQUAFACT International Services Ltd. was commissioned by the Hydraulics and Maritime Research Centre, University College Cork to carry out a marine environmental appraisal of an ocean energy test site in Inner Galway Bay. Figure 1 shows the location of this site. This site has been used to test a wave energy device known as the Ocean Energy Buoy (Figures 2 and 3) since 2006.

This environmental appraisal involved grab sampling five sites within the test site and one control site, approximately 650m south of the test site. Two replicate faunal grab samples were taken at each site in addition to a sediment sample. SCUBA diving was also used to collect sediment profile images and still photography of the sea bed. The report was also to include a desk survey of marine reptiles, mammals and birds that occur in the area and any possible impacts that might arise given the presence of the buoy.



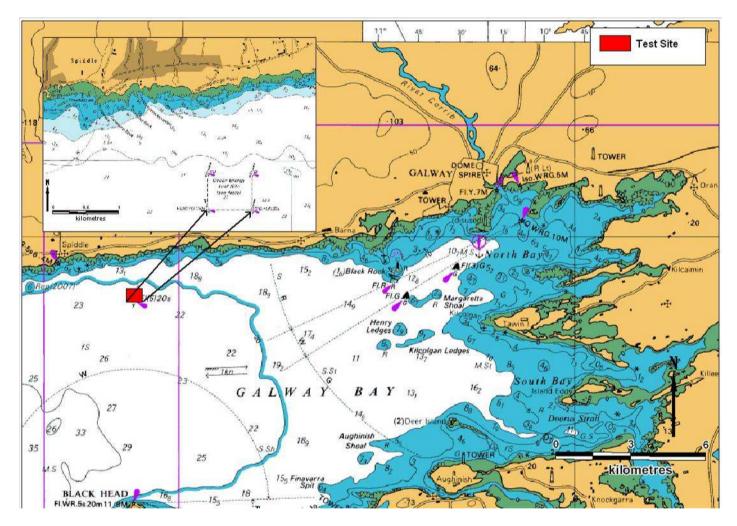


Figure 1: Location of the ocean energy site within Galway Bay.



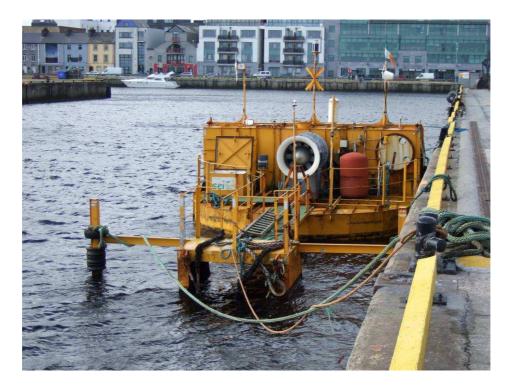


Figure 2: Ocean Energy Buoy moored in Galway Docks.



Figure 3: Ocean Energy Buoy moored in Galway Bay.



2. Marine Benthos

2.1. Sampling Procedure & Processing

To carry out the marine environmental appraisal of the ocean energy test site in inner Galway Bay, AQUAFACT sampled a total of 6 stations. Five of these stations were located within the test site and 1 was located approximately 650m south of the test site and acted as the control station. Figure 4 shows the station locations and Table 1 gives the station coordinates.

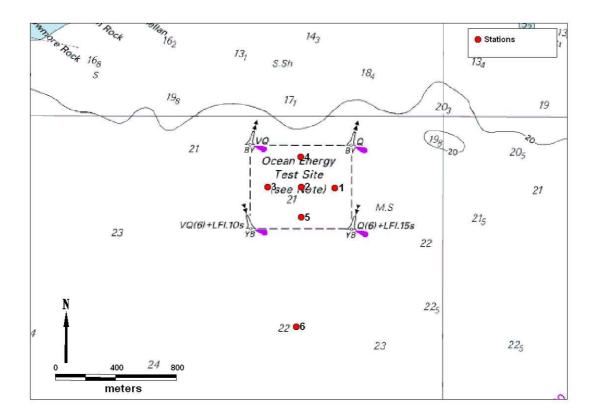


Figure 4: Map showing sampling locations

| Station | Longitude | Latitude | Easting | Northing | Depth (m) |
|---------|-----------|----------|----------|----------|-----------|
| 1 | -9.26079 | 53.2291 | 115851.3 | 220565.6 | 23.2 |
| 3 | -9.26743 | 53.2291 | 115407.9 | 220577 | 22.3 |
| 2 | -9.26411 | 53.2291 | 115630 | 220575.6 | 21.9 |
| 4 | -9.26416 | 53.2309 | 115630 | 220772.8 | 21.3 |
| 5 | -9.26408 | 53.2273 | 115627.9 | 220374.5 | 22.1 |
| 6 | -9.26454 | 53.2208 | 115584.4 | 219647.8 | 25 |

Table 1: Station co-ordinates.

Sampling was carried out on the 7th September 2009 from the *Tarrea Queen*. Stations were located using DGPS and this positioning method is accurate to within c. 1m. A $0.1m^2$ Day Grab was used to collect the benthic samples. Two replicate samples were taken at each of the 6 stations. Data on each sample, e.g. station number, date, time, depth of sediment, surface features and visible macrofauna were logged in a field notebook. The faunal returns were sieved on a 1mm mesh sieve, stained with Rhodamine dye, fixed with 10% buffered formalin and preserved in 70% alcohol. Samples were then sorted under a microscope (x 10 magnification), into four main groups: Polychaeta, Mollusca, Crustacea and others. The 'others' group consisted of echinoderms, nematodes, nemerteans, cnidarians and other lesser phyla. The taxa were then identified to species level where possible.

An additional sample was taken at each station and used for sediment granulometric analyses. The sediment samples were taken through the opening on the top of the grab. All samples were stored immediately in a cold room on board the vessel and were frozen at -20° C on return to the lab.

Particle size analysis was carried out using the traditional granulometric approach by AQUAFACT. Traditional analysis involved the dry sieving of approximately 100g of sediment using a series of Wentworth graded sieves. The process involved the separation of the sediment fractions by passing them through a series of sieves. Each sieve retained a fraction of the sediment, which were later weighed and a percentage of the total was calculated. Table 2 shows the classification of sediment particle size ranges into size classes. Sieves, which corresponded to the range of particle sizes (Table 2) were used in the analysis.



| Range of Particle Size | Classification | Phi Unit |
|------------------------|------------------|--------------------------------|
| <63µm | Silt/Clay | >4 Ø |
| 63-125 μm | Very Fine Sand | 4 Ø, 3.5 Ø |
| 125-250 μm | Fine Sand | 3 Ø, 2.5 Ø |
| 250-500 μm | Medium Sand | 2 Ø, 1.5 Ø |
| 500-1000 μm | Coarse Sand | 1 Ø, 1.5 Ø |
| 1000-2000 μm | Very Coarse Sand | 0 Ø, -0.5 Ø |
| >2000 μm | Gravel | -1 Ø, -1.5 Ø, -2 Ø, -3 Ø, -4 Ø |

| Table 2: The classification of sediment particle size ranges into size classes (adapted from Buchanan, |
|--|
| 1984) |

2.2. Data Processing

All replicates for each station were combined to give a total abundance for each station prior to analyses. A data matrix of all the combined faunal abundance data was compiled and used for statistical analyses. The faunal analysis was carried out using the PRIMER [®] (Plymouth Routines in Multivariate Ecological Research) programme.

Univariate statistics in the form of diversity indices were calculated on the combined replicate data. The following diversity indices were calculated:

1) Margalef's species richness index (D), (Margalef, 1958).

$$D = \frac{S - 1}{\log_2 N}$$

where: N is the number of individuals

S is the number of species

2) Pielou's Evenness index (J), (Pielou, 1977).

$$J = \frac{\dot{H'}(observed)}{\dot{H'}_{max}}$$

where: \dot{H}_{max} is the maximum possible diversity, which could be achieved if all species were equally abundant (= log₂S)

3) Shannon-Wiener diversity index (H'), (Pielou, 1977).

$$H' = -\sum_{i=1}^{s} p_i(\log_2 p_i)$$

where: p_i is the proportion of the total count accounted for by the i^{th} taxa

Species richness is a measure of the total number of species present for a given number of individuals. Evenness is a measure of how evenly the individuals are distributed among different species. The diversity index incorporates both of these parameters. Richness ranges from 0 (low richness) to 12 (high richness), evenness ranges from 0 (low evenness) to 1 (high evenness), diversity ranges from 0 (low diversity) to 5 (high diversity).

The PRIMER [®] manual (Clarke & Warwick, 2001) was used to carry out multivariate analyses on the station-by-station faunal data. It must be noted here that the species that were present only once or twice in the survey were excluded from the multivariate analysis. All species/abundance data were fourth root transformed and used to prepare a Bray-Curtis similarity matrix in PRIMER [®]. The fourth root transformation was used in order to downweigh the importance of the highly abundant species and allow the mid-range and rarer species to play a part in the similarity calculation. The similarity matrix was then used in classification/cluster analysis. The aim of this analysis was to find "natural groupings' of samples, i.e. samples within a group that are more similar to each other, than they are similar to samples in different groups (Clarke & Warwick, *loc. cit.*). The PRIMER [®] programme CLUSTER carried out this analysis by successively fusing the samples into groups and the groups into larger clusters, beginning with the highest mutual similarities then gradually reducing the similarity level at which groups are formed. The result is represented graphically in a dendrogram, the x-axis representing the full set of samples and the y-axis representing similarity levels at which two samples/groups are said to have fused.

The Bray-Curtis similarity matrix was also subjected to a non-metric multi-dimensional scaling (MDS) algorithm (Kruskall & Wish, 1978), using the PRIMER [®] program MDS. This programme produces an ordination, which is a map of the samples in two- or three-dimensions, whereby the placement of samples reflects the similarity of their biological communities rather than their simple geographical location (Clarke & Warwick, 2001). With regard to stress values, they give an indication of how well the multi-dimensional similarity



matrix is represented by the two-dimensional plot. They are calculated by comparing the interpoint distances in the similarity matrix with the corresponding interpoint distances on the 2-d plot. Perfect or near perfect matches are rare in field data, especially in the absence of a single overriding forcing factor such as an organic enrichment gradient. Stress values increase not only with the reducing dimensionality (lack of clear forcing structure), but also with increasing quantity of data (it is a sum of the squares type regression coefficient). Clarke and Warwick (*loc. cit.*) have provided a classification of the reliability of MDS plots based on stress values, having compiled simulation studies of stress value behaviour and archived empirical data. This classification generally holds well for 2-d ordinations of the type used in this study. Their classification is given below:

- Stress value < 0.05: Excellent representation of the data with no prospect of misinterpretation.
- Stress value < 0.10: Good representation, no real prospect of misinterpretation of overall structure, but very fine detail may be misleading in compact subgroups.
- Stress value < 0.20: This provides a useful 2-d picture, but detail may be misinterpreted particularly nearing 0.20.
- Stress value 0.20 to 0.30: This should be viewed with scepticism, particularly in the upper part of the range, and discarded for a small to moderate number of points such as < 50.
- Stress values > 0.30: The data points are close to being randomly distributed in the 2-d ordination and not representative of the underlying similarity matrix.

Each stress value must be interpreted both in terms of its absolute value and the number of data points. In the case of this study, the moderate number of data points indicates that the stress value can be interpreted more or less directly. While the above classification is arbitrary, it does provide a framework that has proved effective in this type of analysis.



2.3. Results

2.3.1. Fauna

The taxonomic identification of the benthic infauna across all 6 stations sampled in the ocean energy test site in inner Galway Bay yielded a total count of 117 species accounting for 1,746 individuals, ascribed to 7 phyla. A complete listing of these species abundance is provided in Appendix 1.

Of the 117 species enumerated, 56 were polychaetes (segmented worms), 31 were crustaceans (crabs, shrimps, prawns), 18 were molluscs (mussels, cockles, snails etc.), 6 species were echinoderms (brittlestars, sea cucumbers), 2 species were cnidarians (sea anemones, corals), 2 species were sipunculids (peanut worms) and 1 species was a phoronid (horseshoe worm).

2.3.1.1. UNIVARIATE ANALYSES

Univariate statistical analyses were carried out on the combined replicate station-by-station faunal data. The following parameters were calculated and can be seen in Table 3; species numbers, number of individuals, richness, evenness and diversity. Species numbers ranged from 49 (S5) to 60 (S1). Number of individuals ranged from 192 (S3) to 389 (S6). Richness ranged from 8.79 (S4) to 10.2 (S2). Evenness ranged from 0.87 (S1) to 0.89 (S3, S4, S5). Diversity ranged from 4.98 (S5) to 5.2 (S6).

| Station | Species | Individuals | Richness | Evenness | Diversity |
|---------|---------|-------------|----------|----------|-----------|
| S1 | 60 | 358 | 10.03 | 0.87 | 5.12 |
| S2 | 59 | 295 | 10.20 | 0.88 | 5.17 |
| S3 | 51 | 192 | 9.51 | 0.89 | 5.08 |
| S4 | 51 | 296 | 8.79 | 0.89 | 5.04 |
| S5 | 49 | 216 | 8.93 | 0.89 | 4.98 |
| S6 | 59 | 389 | 9.73 | 0.88 | 5.20 |

Table 3: Diversity indices for the 6 stations sampled at the ocean energy test site.

2.3.1.2. MULTIVARIATE ANALYSES

The dendrogram and the MDS plot can be seen in Figures 4 and 5 respectively. Two broad groupings can be identified initially. The control station, S6, separated from the other 5 stations at a similarity level of 56.09% (Group I). The remaining stations (all located within the test site) had a similarity level of 60.06%. These stations separated further into Group II and Group III. Group II had a similarity level of 63.15%. This Group consisted of stations, S1 and S4 (65.38% similarity) and station S2 (63.15% similarity). Group III consisted of stations S3 and S5, which had a 60.97% similarity level. It is important to note that all stations were >55% similar to each other indicating a homogeneous faunal community.

Station S6 (Group I) contained 59 species comprising 389 individuals. Of the 59 species present, 38 were present more than twice. Ten species accounted for 50% of the abundance at this station: the polychaetes *Tharyx* sp. (35 individuals; 9%), *Spiophanes kroyeri* (25 individuals; 6.4%), *Chaetozone setosa* (24 individuals; 6.2%), *Lumbrineris gracilis* (21 individuals; 5.4%), *Aricidea (Arcidea) minuta* (19 individuals; 4.9%), *Spiochaetopterus typicus* (17 individuals; 4.4%) *Glycera alba* (15 individuals; 3.9%), the mollusc *Phaxas pellucidus* (15 individuals; 3.9%), the polychaete *Nephtys hombergii* (12 individuals; 3.19%) and the amphipod *Phtisica marina* (12 individuals; 3.1%).

Within Group II, S1 contained 60 species comprising 358 individuals. Of the 60 species present, 33 were present more than twice. Nine species accounted for 50% of the abundance at this station: the polychatetes *Tharyx* sp, (31 individuals; 8.7%). *Glycera alba* (25 individuals; 7%), *Nephtys hombergii* (23 individuals; 6.4%), *Chaetozone setosa* (23 individuals; 6.4%), the mollusc *Thyasira flexuosa* (20 individuals; 5.6%), the polychaetes *Spiochaetopterus typicus* (19 individuals; 5.3%), *Lumbrineris gracilis* (18 individuals; 5%), *Scalibregma inflatum* (15 individuals; 4.2%) and *Ampharete lindstroemi* (15 individuals; 4.2%).

S4, which was 65.38% similar to S1, contained 51 species comprising 296 individuals. Of the 51 species present, 29 were present more than twice. Nine species were responsible for 50% of the abundance: the polychaetes *Scalibregma inflatum* (30 individuals; 10.1%), *Tharyx* sp. (23 individuals; 7.8%), *Spiochaetopterus typicus* (20 individuals; 6.8%), the mollusc *Thyasira*



flexuosa (16 individuals; 5.4%), the amphipod *Ampelisca bre*vicornis (15 individuals; 5.1%), the polychaete *Diplocirrus glaucus* (12 individuals; 4.1%), the mollusc *Phaxas pellucidus* (12 individuals; 4.12%), the polychate *Apistobranchus tullbergi* (11 individuals; 3.7%) and the amphipod *Harpinina serrata* (11 individuals; 3.7%).

S2, which was 63.15% similar to stations S1 and S4, contained 59 species comprising 295 individuals. Of the 59 species present, 30 were present more than twice. Nine species accounted for 50% of the faunal abundance at this station: the amphipod *Photis longicaudata* (35 individuals; 11.9%), the mollusc *Thyasira flexuosa* (18 individuals; 6.1%), the polychaetes *Tharyx* sp. (17 individuals; 5.8%), *Aricidea* (*Arcidea*) minuta (15 individuals; 5.1%), *Ampharete lindstroemi* (15 individuals; 5.1%), *Diplocirrus glaucus* (13 individuals; 4.4%), *Lumbrineris gracilis* (12 individuals; 4.1%), *Spiochaetopterus typicus* (12 individuals; 4.1%) and *Scalibregma inflatum* (10 individuals; 3.4%).

Within Group III, S3 contained 51 species comprising 192 individuals. Of the 51 species present, 19 were present more than twice. Nine species accounted for 50% of the faunal abundance: the polychaetes *Chaetozone setosa* (17 individuals; 8.9%), *Spiochaetopterus typicus* (15 individuals; 7.8%), *Diplocirrus glaucus* (12 individuals; 6.3%), *Ampharete lindstroemi* (11 individuals; 5.7%), the amphipod *Ampelisca brevicornis* (11 individuals; 5.7%), the polychaetes *Tharyx* sp. (10 individuals; 5.2%), *Nephtys hombergii* (9 individuals; 4.7%), *Magelona alleni* (7 individuals; 3.7%)and *Scalibregma inflatum* (7 individuals; 3.7%).

Station S5 which was 60.97% similar to S3 contained 49 species comprising 216 individuals. Of the 49 species present, 24 were present more than twice. Eight species accounted for 50% of the faunal abundance: The molluscs *Thyasira flexuosa* (21 individuals; 9.7%) and *Phaxas pellucidus* (16 individuals; 7.4%), the polychaetes *Tharyx* sp. (15 individuals; 6.9%), *Chaetozone setosa* (14 individuals; 6.5%), *Scalibregma inflatum* (14 individuals; 6.5%), *Spiochaetopterus typicus* (12 individuals; 5.6%), *Diplocirrus glaucus* (12 individuals; 5.6%) and *Lumbrineris gracilis* (7 individuals; 3.2%).

These delineations were also preserved in the MDS plot. The stress value of the MDS ordination is 0.01; this results in an excellent representation of the data with no prospect of misinterpretation.



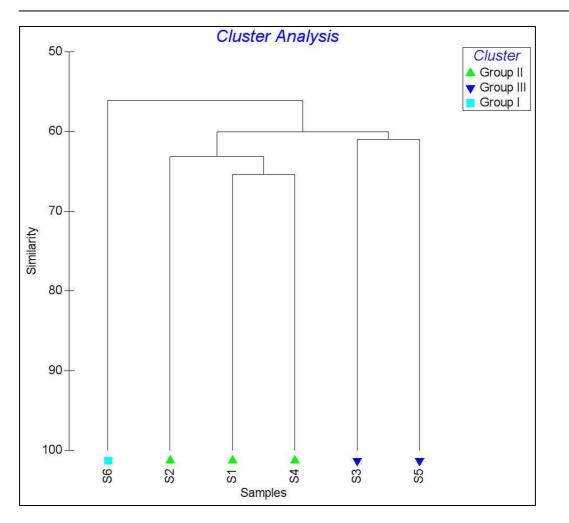


Figure 5: Dendrogram showing each station from the 6 stations sampled in the ocean energy test site.



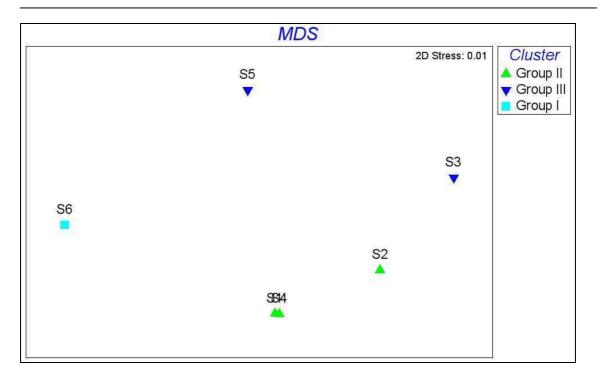


Figure 6: MDS ordination showing each station from the 6 stations sampled in the ocean energy test site.

2.3.2. Sediment

The results from the traditional granulometric analysis can be seen in Table 4. Figure 7 shows this data in graphical form. The sediment sampled during the survey was classified slightly gravelly muddy sand, muddy sand, sand and slightly gravelly sand according to Folk (1954).

The majority of stations were classified as muddy sand (Stations S2, S3 and S4) (See Figure 8). These stations were located in the northwestern part of the test site. Sand was present at station S5 in the southern part of the test site and slightly gravelly muddy sand was present at station S1 in the eastern part of the test site. The control site, S10 was classified as slightly gravelly sand.

Station S1 contained the highest percentage of gravel (1.7%), very coarse sand (1.7%), coarse sand (1.9%), medium sand (2.2%) and silt-clay (13.1%). Station S6 contained the highest percentage of fine sand and station S4 contained the highest percentage of very fine sand (74.5%).



| Station | Gravel* (%) | Very Coarse Sand* (%) | Coarse Sand* (%) | Medium Sand* (%) | Fine Sand* (%) | Very Fine Sand* (%) | Silt- Clay*(%) | Folk (1954)* |
|---------|----------------|-----------------------|---------------------|---------------------|----------------|------------------------|-------------------|---------------------------------|
| 1 | 1.7 | 1.7 | 1.9 | 2.2 | 13.9 | 65.4 | 13.1 | Slightly Gravelly Muddy Sand |
| 2 | 0.3 | 0.7 | 1.2 | 1.6 | 11.8 | 73.9 | 10.6 | Muddy Sand |
| 3 | 0.7 | 1 | 1.5 | 1.8 | 12.6 | 71.4 | 11 | Muddy Sand |
| 4 | 0.2 | 0.7 | 1.3 | 1.7 | 11 | 74.5 | 10.7 | Muddy Sand |
| 5 | 0.5 | 0.9 | 1.4 | 1.8 | 14.4 | 72 | 8.8 | Sand |
| 6 | 1.1 | 0.9 | 1.3 | 1.9 | 24.4 | 63.7 | 6.7 | Slightly Gravelly Sand |

Table 4: Granulometry results for the 6 stations sampled at ocean energy test site in Galway Bay (as percentage weight of the total sample).

*The particle size ranges for each classification (gravel, very coarse sand, coarse sand, medium sand, fine sand, very fine sand and silt clay) is adapted from Buchanan, 1984

and can be seen in Table 2 (page 6). The classifications according to Folk, 1954) are based on varying percentages of gravel, sand and silt-clay.



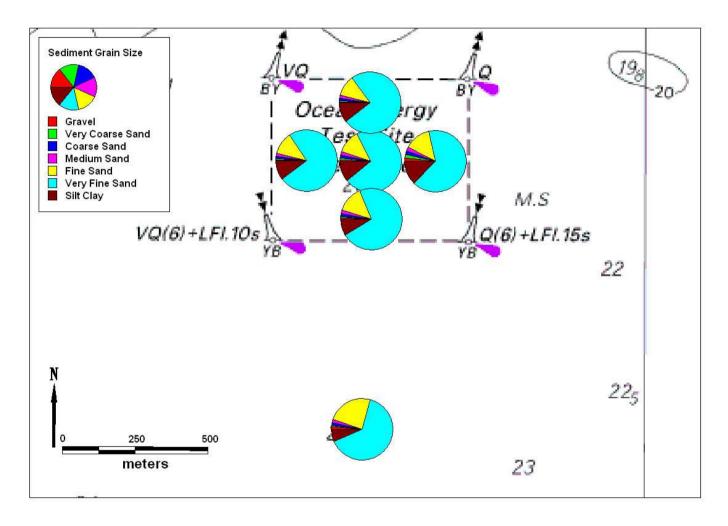


Figure 7: Sediment grain size data



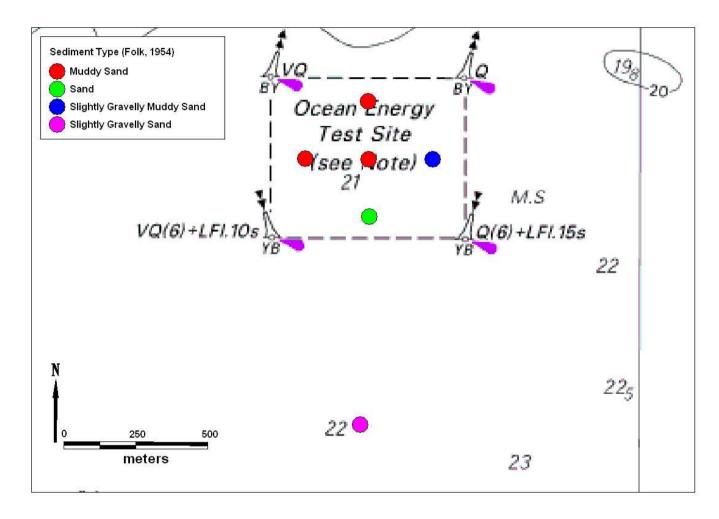


Figure 8: Sediment type according to Folk (1954)



2.3.3. Underwater imaging

Appendix 2 shows a selection of images taken of the sea floor and the mooring chains at the site. The sea bed was found to be a fine, muddy sand with biologically active infauna such as burrowing decapods. Large epibenthic species included *Goneplax rhomboides* and *Marthasterias glacialis*. Mussel shell debris was also noted and these represent material sloughed off the anchor chains and the structure itself. The mooring chains were found to be heavily settled by epifauna such as anemones, mussels, star fish and echinoids.

2.3.3.1. Sediment Profile Imaging (SPI)

SPI images were taken within the location where the ocean energy device had been deployed and were uniform in many respects (see Appendix 3 for representative photographs). The sediment was found to be a fine, muddy sand with well bioturbated sediments giving rise to deep redox layers throughout the site. Where present, surface boundary roughness was biologically-generated. These images indicated a biologically active benthos with a Stage III community present.

2.3.4. Discussion

The sediment type within the test site varied between muddy sand, sand and slightly gravelly muddy sand. The control site, station S6, was classified as slightly gravelly sand.

Species richness is a measure of the total number of species present for a given number of individuals. Evenness is a measure of how evenly the individuals are distributed among different species. The diversity index incorporates both of these parameters. Richness ranges from 0 (low richness) to 12 (high richness), evenness ranges from 0 (low evenness) to 1 (high evenness), diversity ranges from 0 (low diversity) to 5 (high diversity).

The results from the univariate statistical analyses are all typical of muddy coastal sediments off the West coast.



| Station | Species | Individuals | Richness | Evenness | Diversity |
|---------|---------|-------------|----------|----------|-----------|
| S1 | 60 | 358 | 10.03 | 0.87 | 5.12 |
| S2 | 59 | 295 | 10.20 | 0.88 | 5.17 |
| S3 | 51 | 192 | 9.51 | 0.89 | 5.08 |
| S4 | 51 | 296 | 8.79 | 0.89 | 5.04 |
| S5 | 49 | 216 | 8.93 | 0.89 | 4.98 |
| S6 | 59 | 389 | 9.73 | 0.88 | 5.20 |

| Table E: Number of Sr | nacios Individual | thair Dichnose | Evenness and Diversity | , at each Station |
|-----------------------|---------------------|--------------------|--------------------------|-------------------|
| rable 5: Number of Sp | pecies, individuals | s, their Richness, | , Evenness and Diversity | at each Station |

A benthic faunal study carried out by O'Connor & McGrath (1981) in inner Galway Bay in 1976 characterised the ocean energy test site as an *Abra nitida/Turritella communis* variation of the *Cylichna cylindracea* and *Ampelisca spinipes* silty sand community. The stations sampled in the present study revealed elements of the *Cylichna cylindracea* and *Ampelisca spinipes* silty sand community, however the *Abra nitida/Turritella communis* variation of this community was not evident.

The dominant species from this survey were the polychaetes *Tharyx* sp., *Scalibregma inflatum*, *Spiophanes kroyeri*, *Glycera alba*, *Spiochaetopterus typicus*, *Chaetozone setosa*, the amphipod *Photis longicaudata* and the molluscs *Thyasira flexuosa* and *Phaxas pellucidus*. Species richness and evenness was high at each station.

The results of the multivariate analyses indicate a high level of similarity between the sampled locations and suggest that there has been no noticeable change in benthic fauna within the site. This is further supported by the high level of similarity between the "in site" data and the control.

The SPI images indicate a healthy Stage community present at the site with both infaunal and epibenthic (decapods) species reworking and oxygenating the upper *ca* 15 cms of sediment to such a degree that no discontinuity in oxygen levels were noted on any of the images collected.

The only noticeable difference between the test site samples and the control was the



occasional occurrence of mussel (*Mytilus edulis*) shells within the test area. These shells were probably sloughed off the ocean energy device while *in situ* or when it was being moved into the docks at Galway.

3. Marine reptiles, mammals and birds

3.1. Marine reptiles of Ireland's Atlantic margin.

Five species of marine turtle have been recorded in UK and Irish waters (Brongersma, 1972; Penhallurick, 1990; Langton *et al.*, 1996; Gaywood, 1997; Pierpoint & Penrose, 1999). Only one species however, the leatherback turtle *Dermochelys coriacea* is reported annually and is considered a regular and normal member of Irish marine fauna (Godley *et al.* 1998). Loggerhead turtles *Caretta caretta* and Kemp's ridley turtles *Lepidochelys kempii* occur less frequently, with most specimens thought to have been carried north from their usual habitats by adverse currents (Carr, 1987; Penhallurick, 1990; Mallinson, 1991). Records of two other vagrant species, the hawksbill turtle *Eretmochelys imbricata* and the green turtle *Chelonia mydas* are very rare (Brongersma, 1972; O'Riordan *et al.*, 1984; Branson, 1997). Due to the rarity of the latter 4 species in the ocean energy device area and wider Galway Bay area, they are not discussed further.

Leatherback turtle (Dermochelys coriacea)

The leatherback turtle is the largest marine turtle that occurs in warm waters and has been regularly recorded in Irish waters. They breed circumglobally within latitudes approximately 40°N and 35°S, but range widely to forage in temperate and boreal waters outside the nesting season (Eckert, 1995). They are the only species of marine turtle to have developed adaptations to life in cold water (Greer *et al.* 1973; Goff & Stenson, 1988). It is an endangered species throughout its distributional range. The total number of leatherbacks nesting world-wide in 1995 was estimated at 34,529 (confidence interval 26,177 to 42,878) females (Spotila *et al.*, 1996). About 80% of these animals were reported from sites in the Atlantic. Long-distance migration has been documented from tag returns and more recently using satellite telemetry. There are distinct seasonal peaks in the occurrence of leatherback turtles in northern waters. Around the UK, most turtles are reported between August and

October (Gaywood, 1997; Godley *et al.*, 1998). Using an expanded dataset, Pierpoint & Penrose (1999) report that leatherbacks have been reported from UK and Irish waters in every month, although live sightings peak in August. Strandings peak slightly later, in September and October. Leatherback turtles feed primarily on jellyfish. Their diet in temperate and boreal waters is known to include cnidarians (jellyfish) and tunicates (seasquirts) (salps, pyrosomes) (den Hartog & van Nierop, 1984; Davenport & Balazs, 1991). In UK and Irish waters they are often reported in the vicinity of jellyfish swarms, and there are several observations of leatherbacks feeding on jellyfish at the surface (e.g. Brongersma, 1972; Penhallurick, 1990). The leatherback turtle is included in the EU Habitats Directive and CITES (Convention on International Trade in Endangered Species of Wild Fauna and Flora). This species has been recorded in the Galway/Clare area and may be an occasional visitor to the ocean energy device site. As it is generally on the sea surface, it is considered unlikely that it would be affected by the workings of the device.

3.2. Marine Mammals of Ireland's Atlantic Margin

Irish waters are some of the most important in Europe for a wide range of cetacean (whales, dolphins and porpoise) species (Berrow, 2001). There is substantial background evidence promoting the Atlantic Margin as an area of high species richness for cetaceans (Ó Cadhla *et al.*, 2004). To date, 24 cetacean species have been recorded in Irish waters (Berrow, 2001). The majority of these have been recorded from sightings or acoustic recordings (as well as from strandings), whereas only three species have only been recorded as strandings, they are Gervais's beaked whale, pygmy sperm whale and True's beaked whale (IWDG, *pers. comm.*). Three species of pinnipeds (seals) have been reported from Irish waters.

Parturition (breeding) in Irish waters has been confirmed for a number of cetacean species including harbour porpoise *Phocoena phocoena*, common *Delphinus delphis*, bottlenose *Tursiops truncatus*, Risso's *Grampus griseus*, white sided *Lagenorhynchus acutus* and white-beaked *L albirostris* dolphins and pilot whale *Globiocephala melas*, while other species such as bottlenosed *Hyperoodon ampullatus* and minke whale *Balaenoptera acutorostrata* are also suspected of breeding. Many are not known to breed in Irish waters but migrate annually along the western seaboard (Charif & Clark, 2000), these include the blue whale *Balaenoptera musculus*, fin whale *B. physalus* and humpback whale *Megaptera novaegliana*.



Recent data suggests that some of these species feed year-round in waters along the south coast, including the fin and humpback whale (IWDG, *pers. comm.*), whereas others may over-winter in waters south of Ireland e.g. blue whale *Balaenoptera musculus* (Charif & Clark, 2000). Others such as beluga *Delphinapterus leucas* are vagrants on the edge of their range in the Northeast Atlantic and only occur infrequently

The grey and harbour seals are the two seal species native to Irish waters. Both species have established themselves in terrestrial colonies (or haul-outs) along all coastlines of Ireland, which they leave when foraging or moving between areas and to which they return to rest ashore, rear young, engage in social activity, etc. The haul-out groups of harbour seals have tended historically to be found among inshore bays and islands, coves and estuaries (Lockley, 1966; Summers *et al.*, 1980), particularly around the hours of lowest tide. The grey seal breeds on exposed rocky shores, on sand bars or sea caves with ready access to deep water. Other haul-out areas for the grey seal are located on exposed rocky areas or steeply shelving sandbanks. Fur seal and walrus are also visitors to Atlantic waters (Ó Cadhla, *pers. comm.*).

Figure 9 below shows all whales and dolphins recorded from Galway Bay from January 2008 (Source: IWDG Sightings).



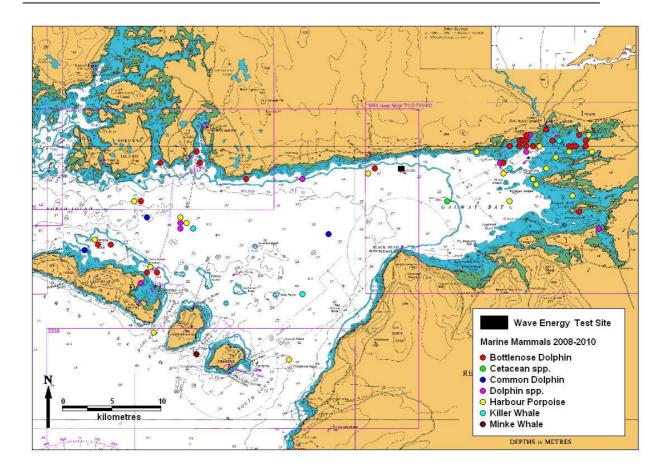


Figure 9: Map showing Wave energy test site and areas where cetaceans were recorded between 2008 and 2010

All marine mammals are considered to be able to swim at fast enough velocities so as to be able to avoid the physical actions of the device.

3.3. Birds of Ireland's Atlantic Margin

The exposed and inaccessible west coast of Ireland provides a perfect breeding habitat for many seabird species. In addition to this, coastal and offshore Irish waters provide local breeding and non-breeding seabirds, along with pelagic and passage migrants, with a rich source of nutrition, particularly near coastal upwelling and frontal systems (e.g. along the Irish Shelf front, north of the Porcupine Seabight). The availability and distribution of prey are considered to be the most important factors driving seabird distribution and abundance (Skov *et al.*, 1995; Begg & Reid, 1997; Ollason *et al.*, 1997), particularly around colonies



during the breeding season (Boelens, *et al.,* 1999). The waters off western and southern Ireland are also internationally important fishing grounds and important nursery and spawning areas for fish and invertebrate species. Areas off the west coast of Ireland are regarded to be of the highest importance as feeding areas for seabirds.

Mackey *et al.* (2004) described the distribution, density and relative abundance of seabirds of Ireland's Atlantic Margin. The Atlantic Margin is characterised by a number of physical, climate and oceanographic features, which combine to produce one of the most productive areas of the northeast Atlantic Ocean. As a result, the waters off western and southern Ireland contain internationally renowned commercial fishing grounds (e.g. Porcupine Bank, Celtic Sea), important nursery and spawning areas for fish and invertebrate species, and areas of concentration for foraging seabirds and other predators (Boelens *et al.*, 1999).

3.3.1. Inner Galway Bay

Inner Galway Bay Survey

Inner Galway Bay is an internationally important bird site. The habitat of the site is composed of open rocky/boulder/shingle coastline along the northern and southern shores, estuarine habitat with shallow sandy bays and lagoons between Oranmore and Kinvarra, shallow tidal inlet bordered by saltmarsh (Lough Atalia) and an adjacent turlough (Ahapouleen). This site has been regularly monitored since the *Wetlands Enquiry (1971-74)*. It is currently counted in four parts, all of which have been regularly monitored during all I-WeBS seasons. The total count area of the site is 10,321 hectares (103.21 km2). In more recent seasons (since the period of the Crowe, 2005 publication), these four sections have been further sub-divided into 24 count units.

Table 6 shows the mean number of water birds present in the area from 1996 to 2000.

| Group | Mean No. |
|---------------------------------------|----------|
| Number of regularly occurring species | 40 |
| Wildfowl | 4,424 |
| Waders | 11,544 |
| Gulls | 3,386 |



Inner Galway Bay supports the greatest diversity of waterbirds in the country. It is internationally important for great northern diver and light-bellied Brent goose and nationally important for a further 15 species. This site also supports significant concentrations of both black-headed and common gulls. Waterbird numbers overall have increased (by 51%) since the *Winter Wetlands (1984-86)*. There have been increases in most species, particularly great northern diver, cormorant, shoveler and wader species such as golden plover, lapwing, dunlin and bar-tailed godwit. The majority of species have continued to increase or remain stable at Inner Galway Bay during the I-WeBS period. Overall, the difference between the first five season peak mean (1994/95 to 1998/99) and the most recent (to 2000/01) illustrates that both wildfowl and waders have increased by 14% and 25% respectively. The long-term species declines in Nairn *et al.* (2000), such as grey heron, ringed plover, redshank and turnstone have either stabilised or reversed during I-WeBS.

The northeast and east sections of the bay generally support the majority of birds, while the islands of the inner bay are used by large concentrations of roosting waders. The open water habitats are of particular importance for great northern diver, little grebe, cormorant and red-breasted merganser. Numbers of some species vary considerably. Golden plover, lapwing and curlew regularly move inland to feed on fields, returning to the shoreline in large numbers when the ground is frozen. Some divers, seaducks and gulls are occasionally located far offshore and out of observation range, particularly when sea conditions are rough. However, this site is regularly counted, thereby increasing the likelihood that these flocks are counted at some stage in a given season.

Table 7 shows the significant concentrations of waterbirds in Inner Galway Bay between 1994/94 and 2000/01.



Table 7: Concentrations of waterbirds in Inner Galway Bay between 1994/95 and 2000/01.

| Species | 94-98 | 95-99 | 96-00 | Month | % Flyway | % | Rank | % change |
|----------------------|-------|-------|-------|--------|----------|----------|------|----------|
| | | | | | | National | | (WWS) |
| International | I | 1 | | • | | | | |
| Great Northern Diver | 78 | 83 | 105 | Mar | 2.1 | 5.3 | 1 | +401 |
| Light-bellied Brent | 638 | 682 | 746 | Mar | 3.7 | 3.7 | 9 | +39 |
| Goose | | | | | | | | |
| National | 1 | 1 | 1 | | | | | |
| Cormorant | 249 | 266 | 285 | Nov | 0.2 | 1.9 | 7 | +65 |
| Shelduck | 127 | 146 | 155 | Jan | 0.1 | 1.0 | 23 | +24 |
| Wigeon | 1,150 | 1,168 | 1,187 | Jan | 0.1 | 1.3 | 23 | +2 |
| Teal | 612 | 700 | 797 | Jan | 0.2 | 1.4 | 19 | +22 |
| Shoveler | 96 | 88 | 130 | Nov/Ja | 0.3 | 3.3 | 8 | +66 |
| | | | | n | | | | |
| Red-breasted | 250 | 249 | 250 | Nov | 0.2 | 6.3 | 1 | +17 |
| Merganser | | | | | | | | |
| Ringed Plover | 297 | 335 | 287 | Jan | 0.4 | 1.9 | 5 | -47 |



Ocean Energy Test Site in Inner Galway Bay

April 2010

| Species | 94-98 | 95-99 | 96 | -00 | Month | | % Fly | way | % | | Rank | % | change | |
|-------------------|-------|-------|-------|-------|------------|--------|-------|-----|----------|----|-------|-----|--------|--|
| | | | | | | | | | National | | | (ww | /S) | |
| Golden Plover | 1,584 | 2,030 | 2,1 | 2,171 | | /M | 0.2 | | 1.5 | | 35 | +98 | | |
| | | | | | ar | | | | | | | | | |
| Lapwing | 3,360 | 3,969 | 4,1 | 189 | Jan | | 0.2 | | 2.1 | | 13 | +18 | +18 | |
| Dunlin | 1,642 | 2,155 | 2.2 | 203 | Jan | | 0.2 | | 1.6 | | 14 | +75 | +75 | |
| Bar-tailed Godwit | 402 | 447 | 47 | 475 | | 0.4 | | | 2.6 | | 12 | +75 | +75 | |
| Curlew | 650 | 697 | 689 | | Nov/Ja 0.2 | | | 1.0 | | 21 | +36 | +36 | | |
| | | | | | | n | | | | | | | | |
| Redshank | 506 | 505 | 50 | 8 | Nov | | 0.3 | | 1.5 | | 18 | -20 | | |
| Greenshank | 20 | 20 | 18 | | Nov | | ~ | | 0.9 | | 18 | -4 | | |
| Turnstone | 160 | 182 | 22 | 23 | Nov | | 0.2 | | 1.6 | | 7 -15 | | | |
| Gulls | | | | | | | | | | | | | | |
| Black-headed Gull | 1,571 | 1,815 | | 2,108 | |)8 Jan | | ~ | | - | | 14 | | |
| Common Gull | 988 | 1,017 | 017 9 | | | Jan | | 0.1 | | - | | 7 | | |



| Inner Galway Bay | One of the most important ornithological sites in the western region. |
|------------------|--|
| (004031) | Supports an excellent diversity of wintering wetland birds, with divers, |
| | grebes, cormorants, dabbling duck, sea duck and waders all well |
| | represented. Internationally important wintering populations of Great |
| | Northern Diver (83) and Brent Goose (676). Nationally important |
| | populations of an additional sixteen species, i.e. Black-throated Diver |
| | (25), Cormorant (266), Mute Swan (150), Wigeon (1,157), Teal (690), |
| | Shoveler (88), Red-breasted Merganser (249), Ringed Plover (335), |
| | Golden Plover (2,030), Lapwing (3,969), Dunlin (2,149), Bar-tailed |
| | Godwit (447), Curlew (697), Redshank (505), Greenshank (20) & |
| | Turnstone (182) – all figures are average peaks for the 5 seasons |
| | 1995/96-1999/00. Of note is that the populations of Red-breasted |
| | Merganser and Ringed Plover represent 6.7% and 3.3% of the |
| | respective national totals. Black-throated Diver is a scarce species in |
| | Ireland and the Galway Bay population is the most regular in the |
| | country. Other species which occur in notable numbers include Little |
| | Grebe (35), Grey Heron (102), Longtailed Duck (19) and Scaup (40). The |
| | bay is an important wintering site for gulls, especially Black-headed Gull |
| | (1,815), Common Gull (1,011) and Herring Gull (216). In addition, the |
| | following species also use the site: Red-throated Diver (13), Great |
| | Crested Grebe (16), Mallard (200), Shelduck (139), Common Scoter (79), |
| | Oystercatcher (575), Grey Plover (60), Black-tailed Godwit (45) and |
| | Great Blackbacked Gull (124). The site provides both feeding and roost |
| | sites for most of the species, though some birds also commute to areas |
| | outside of the site. The site has several important populations of |
| | breeding birds, most notably colonies of Sandwich Tern (81 pairs in |
| | 1995) & Common Tern (99 pairs in 1995). A large Cormorant colony |
| | occurs on Deer Island (300 pairs in 1989). |

Of the above species, the only ones that might occur at the wave energy site would be Great Northern, Black throated and Red Throated Diver, cormorant, shag, red breasted merganser, Gull and tern species. Other species that will occur on the site include Manx, Sooty, Great and Cory's shearwater, storm and fulmar petrels, Arctic and Great skua, gannet, white



backed gull species, puffin, black and common guillemot and razorbill. There is also a number of rarer species such as Wilson's petrel, Little and Sabine's gull, little auk and grey phalarope that most likely can occur at the site. Eider duck are now known to breed at Slyne Head and if this southward extension of their distribution continues, this species may move through the ocean energy site with time.

Regarding these species, only the diving birds can be affected by ocean energy devices. These species are sooty shearwater, Great Northern, Black throated and Red Throated Diver, cormorant, shag, red breasted merganser, tern and auk species. The ocean energy device draws in a column of water that is ca 1 m in height and this same volume of water is exhaled approximately 6 seconds later. Any bird that may be drawn into the device will therefore be flushed out with this water.

Observations of the device show that gulls use it as a roosting and feeding site. Gulls were seen to alight on the structure and stay there for periods of time and were also seen to collect epifaunal species such as mussels and feed on them on the device.

4. References

Begg, G.S & J.B. Reid. 1997. Spatial variation in seabird density at a shallow sea tidal mixing front in the Irish Sea. *ICES J. Mar. Sci.* **54**: 552-565.

Berrow, S. 2001. Biological diversity of cetaceans (whales, dolphins and porpoises) in Irish waters. In: J.D. Nunn (ed.), *Marine Biodiversity in Ireland and adjacent waters*. Ulster Museum, Belfast. pp. 115-120.

Boelens, R.G.V., Maloney, D.M., Parsons, A.P. & A.R. Walsh. 1999. *Ireland's marine and coastal areas and adjacent seas. An environmental assessment*. Marine Institute. Dublin. 388 pp.

Branson, A. 1997. Cetaceans and sea life. British Wildlife 8(4): 250.

Brongersma, L.D. 1972. European Sea Turtles. Zoologische Verhandelingen - uitgegeven door het rijksmuseum van natuurlijke te Leiden, Nr. 121, Leiden, E.J. Brill

Buchanan, J.B. (1984). Sediment analysis. In: (eds.) Holme N. A. and A.D. McIntyre. Methods for the study of marine benthos 2nd ed. *Blackwell, Oxford.* pp. 41-65.



Carr, A.F. 1987. New perspectives on the pelagic stage of marine turtle development. *Cons. Biol.* **1(2):** 103-121.

Clarke, K.R. and R.M. Warwick (2001). Changes in marine communities: An approach to statistical analysis and interpretation. 2nd Edition. *Primer-E Ltd.*

Charif, R.A. & C.W. Clark. 2000. Acoustic monitoring of large whales to the west of Britain and Ireland using bottom mounted hydrophone arrays, October 1996-September 1998. JNCC report, no. 313.

Crowe, O. 2005. Ireland's wetlands and their waterbirds: status and distribution. Birdwatch Ireland publication. pp 402.

Davenport, J. & G.H. Balazs. 1991. 'Fiery bodies' – are pyrosomas an important component part of the diet of Leatherback turtles? *Brit. Herpetol. Soc. Bull.* **31**:33-38.

Eckert K L. 1995. Leatherback sea turtle, Dermochelys coriacea. In: P.T. Plotkin National Marine Fisheries Service and US Fish and Wildlife Service status reviews for sea turtles listed under the Endangered Species Silver Spring, Maryland, USA, National Marine Fisheries Service. pp. 37-75.

Folk, R.L. (1954). The distinction between grain size and mineral composition in sedimentary rock nomenclature. *Journal of Geology* **62 (4):** 344-359.

Gaywood, M.J. 1997. Marine turtles in British and Irish waters. British Wildlife 9(2): 69-77

Godley, B., Gaywood, M., Law, R., McCarthy, C., McKenzie, C., Patterson, I., Penrose, R., Reid, R. & H. Ross. 1998. Patterns of marine turtle mortality in British waters 1992-96 with reference to tissue contaminant levels. *Journal of the Marine Biological Association UK* **78**: 973-984

Goff, G.P. & G.B. Stenson. 1988. Brown adipose tissue in Leatherback sea turtles: a thermogenic organ in an endothermic reptile? *Copeia* 1071-1075.

Greer, A.E., Lazell, J.D. & R.M. Wright. 1973. Anatomical evidence for counter-current heat exchanger in the Leatherback turtle (*Dermochelys coriacea*). *Nature* 244: 181

Hartog, J.C. den & M.M. van Nierop. 1984. A study of the gut contents of six Leathery turtles *Dermochelys coriacea* (Linnaeus) (Reptilia: Tesudines: Dermochelydae) from British waters and from the Netherlands. *Zoologische Verhandlingen*, 200. Leiden, Rijksmuseum van Natuurlijke Historie

Kruskall, J.B. % M. Wish (1978). Multidimensional scaling. Sage Publications, Beverly Hills, California

Langton, T.E.S, Beckett, C.L., King, G.L. & M.J. Gaywood. 1996. Distribution and status of

marine turtles in Scottish waters. Scottish Natural Heritage Research, Survey and Monitoring Report, No. 8.

Lockley, R.M. 1966. The distribution of Grey and Common seals on the coasts of Ireland. *Irish Naturalists' Journal* **15:** 136–43.

Mackey, M., O' Cadhla, O., Kelly, T.C., Aguilar de Soto, N. & N. Connolly. 2004. *Cetaceans and Seabirds of Ireland's Atlantic Margin. Volume I – Seabird distribution, density & abundance.* Report on research carried out under the Irish Infrastructure Programme (PIP): Rockall Studies Group (RSG) projects 98/6 and 00/13, Porcupine Studies Group project P00/15 and Offshore Support Group (OSG) project 99/38. 95 pp.

Mallinson, J.J. 1991. Stranded juvenile Loggerheads in the United Kingdom. *Marine Turtle Newsletter* **54:** 14-16.

Margalef, DR. (1958). Information theory in ecology. *General Systems* **3**: 36-71.

Nairn, R.G.W., ten Carte, M.E. & N. Sharkey. 2000. Long term monitoring of wintering waterbirds in inner Galway Bay, 1980/1981 to 1999/2000. *Irish Birds* **6**: 453-468.

Ó Cadhla, O., Mackey, M., Auguilar de Soto, N., Rogan, E. & N. Connolly. 2004. *Cetaceans and Seabirds of Ireland's Atlantic Margin. Volume II – Cetacean distribution and abundance.* Report on research carried out under the Irish Infrastructure Programme (PIP): Rockall Studies Group (RSG) projects 98/6 and 00/13, Porcupine Studies Group project P00/15 and Offshore Support Group (OSG) project 99/38. 82pp.

O'Connor, B & D. McGrath. 1981. Benthic macrofaunal studies in the Galway Bay area,

Volume I: The Macrobenthic Faunal Assemblages of Galway Bay. Ph. D Thesis, University College Galway.

Ollason, J.G., Bryant, A.D., Davis, P.M., Scott, B.E. & M.L. Tasker. 1997. Predicted seabird distributions in the North Sea: the consequences of being hungry. *Ices J. Mar. Sci.* **54**: 505-517.

O'Riordan, C.E., Holmes, J.M.C. & D.P. Sleeman. 1984. First recorded occurrence of the Hawksbill Turtle (*Eretmochelys imbricata* (L)) in Irish waters. *Irish Naturalists Journal* **21(6)**: 274-5.

Penhallurick, R.D. 1990. Turtles off Cornwall, the Isles of Scilly and Devonshire. Truro, Cornwall, Dyllansow Pengwella, ISBN 0-9515785-0-2.

Pielou, E.C. (1977). Mathematical ecology. Wiley-Water science Publication, John Wiley and Sons. pp.385.

Pierpoint, C. & Penrose. 1999. TURTLE: A Database of Marine Turtle Records for the United

Kingdom & Eire, Version 1 (Oct. 1999): Introduction, Data Summary & User Notes. (Contractor: Marine Environmental Monitoring, Llechryd.) Unpublished report to English Nature

Skov, H., Durinck, J., Leopold, M.F. & M.L. Tasker. 1995. *Important bird areas for seabirds in the North Sea.* Birdlife International. Cambridge. 156 pp.

Spotila, J.R., Dunham, A.E., Leslie, A.J., Steyermark, A., Plotlin, P.T. & F.V. Paladino. 1996. Worldwide population decline of *Dermochelys coriacea*: are Leatherback turtles going extinct? *Chelonian Conservation and Biology* **2**: 209-222.

Summers, C.F., Warner, P.J., Nairn, R.G.W., Curry, M.G. & J. Flynn. 1980. An assessment of the status of the Common seal *Phoca vitulina* in Ireland. *Biological Conservation* **17 (2):** 115-123.



Appendix 1 Faunal Abundance List

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|------------------------|---|-----|----|----|----|----|----|----|----|----|----|----|----|----|
| CNIDARIA | D | 1 | | | | | | | | | | | | |
| ACTINIARIA | D | 662 | | | | | | | | | | | | |
| Haloclavidae | D | 749 | | | | | | | | | | | | |
| Peachia hastata | D | 755 | | 2 | | | 1 | | | | | | | |
| Edwardsiidae | D | 759 | | | | | | | | | | | | |
| Edwardsia sp. | D | 764 | 4 | | | | | | | | | | | |
| SIPUNCULA | Ν | 1 | | | | | | | | | | | | |
| SIPUNCULIDEA | Ν | 2 | | | | | | | | | | | | |
| GOLFINGIIFORMES | Ν | 10 | | | | | | | | | | | | |
| Golfingiidae | Ν | 11 | | | | | | | | | | | | |
| Nephasoma | Ν | 25 | | 3 | | | | 3 | | | 1 | | 3 | 1 |
| (Nephasoma) minutum | | | | | | | | | | | | | | |
| Phascolionidae | Ν | 29 | | | | | | | | | | | | |
| Phascolion strombus | Ν | 37 | | | 1 | | 1 | | | | | | | |
| ANNELIDA | Р | 1 | | | | | | | | | | | | |
| POLYCHAETA | Р | 2 | | | | | | | | | | | | |
| PHYLLODOCIDA | Р | 3 | | | | | | | | | | | | |
| Polynoidae | Р | 25 | | | | | | | | | | | | |
| Malmgreniella | Р | | | | | 1 | | | 1 | 1 | | | | |
| arenicolae | | | | | | | | | | | | | | |
| Pholoidae | Р | 90 | | | | | | | | | | | | |
| Pholoe inornata | Р | 92 | | | | | | 2 | | | 2 | | | |
| Pholoe synophthalmica | Р | 94 | 9 | | | 2 | | | | | | | | |
| Sigalionidae | Р | 96 | | | | | | | | | | | | |
| Sthenelais sp. | Р | 106 | | | | | | | | | 1 | | | |
| Sthenelais cf limicola | Р | 109 | 2 | | 1 | 2 | 3 | 1 | 2 | | | 2 | 2 | |
| Phyllodocidae | Р | 114 | | | | | | | | | | | | |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|-------------------------|---|-----|----|----|----|----|----|----|----|----|----|----|----|----|
| Eteone flava | Р | 117 | | | | | | 2 | | | | | | |
| Phyllodoce (Anaitides) | Р | 141 | | | | 1 | | | | | | | | |
| groenlandica | | | | | | | | | | | | | | |
| Phyllodoce longipes | Р | 143 | | 1 | | | | | | | | | | |
| Phyllodoce (Anaitides) | Р | 148 | | 1 | | | | | | | | | 1 | |
| rosea | | | | | | | | | | | | | | |
| Eumida bahusiensis | Р | 164 | | 3 | 2 | 5 | | | | | | 1 | 8 | 3 |
| Paranaitis kosteriensis | Р | 176 | | | 2 | | | 2 | | | | | | |
| Glyceridae | Р | 254 | | | | | | | | | | | | |
| Glycera alba | Р | 256 | 14 | 11 | 3 | | 2 | | 7 | 3 | 2 | 3 | 13 | 2 |
| Glycera rouxii | Р | 263 | | | 3 | | | 2 | | | | | | |
| Goniadidae | Р | 266 | | | | | | | | | | | | |
| Goniada maculata | Р | 271 | | | 2 | | | | | | 1 | | | |
| Hesionidae | Р | 293 | | | | | | | | | | | | |
| Ophiodromus flexuosus | Р | 313 | | | | 2 | | 1 | | 1 | 3 | 3 | 4 | |
| Podarkeopsis capensis | Р | 319 | | | | 2 | | | | | | | | |
| Syllidae | Р | 346 | | | | | | | | | | | | |
| Exogone hebes | Р | 421 | | | | | | | 2 | | | | | |
| Autolytus sp. | Р | 434 | | | | | | | | | 1 | | | |
| Nephtyidae | Р | 490 | | | | | | | | | | | | |
| Nephtys sp. (juv) | Р | 494 | | | | | | | | | | | | 3 |
| Nephtys cirrosa | Р | 498 | 1 | | | | | | | | | | | |
| Nephtys hombergii | Р | 499 | 9 | 14 | 4 | 3 | 5 | 4 | 3 | 4 | | | 12 | |
| EUNICIDA | Р | 536 | | | | | | | | | | | | |
| Lumbrineridae | Р | 569 | | | | | | | | | | | | |
| Lumbrineris gracilis | Р | 579 | 11 | 7 | 6 | 6 | 3 | 3 | 7 | | 4 | 3 | 14 | 7 |
| Abyssoninoe hibernica | Р | | | | | | | | | | | 1 | 2 | |
| Dorvilleidae | Р | 598 | | | | | | | | | | | | |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|--------------------------|---|-----|----|----|----|----|----|----|----|----|----|----|----|----|
| Protodorvillea | Р | 638 | | | | | | | | 1 | | | | |
| kefersteini | | | | | | | | | | | | | | |
| ORBINIIDA | Р | 654 | | | | | | | | | | | | |
| Orbiniidae | Р | 655 | | | | | | | | | | | | |
| Scoloplos armiger | Р | 672 | 3 | 1 | | | | | | | | 1 | | 3 |
| Paraonidae | Р | 674 | | | | | | | | | | | | |
| Aricidea (Arcidea) | Р | 677 | 7 | | 8 | 7 | 2 | 2 | 4 | 2 | | 7 | 19 | |
| minuta | | | | | | | | | | | | | | |
| Paradoneis lyra | Р | 699 | | 1 | | | | | | | | | | 3 |
| SPIONIDA | Р | 707 | | | | | | | | | | | | |
| Apistobranchidae | Р | 709 | | | | | | | | | | | | |
| Apistobranchus tullbergi | Р | 712 | 2 | | 1 | | | | 2 | 9 | | 3 | 4 | 3 |
| Spionidae | Р | 720 | | | | | | | | | | | | |
| Prionospio sp. | Р | 763 | | | | 1 | 3 | | | | 4 | | 4 | 7 |
| Prionospio fallax | Р | 765 | | | | | | | | | | | 1 | |
| Spio filicornis | Р | 790 | | | | | 3 | | | | 2 | 2 | | |
| Spiophanes bombyx | Р | 794 | 2 | 2 | | 2 | | | 3 | | | | | |
| Spiophanes kroyeri | Р | 796 | | | | | | 2 | | 2 | 3 | | 16 | 9 |
| Magelonidae | Р | 802 | | | | | | | | | | | | |
| Magelona sp. | Р | 803 | | | | | | | | | | | 1 | |
| Magelona alleni | Р | 804 | | 2 | 2 | 3 | 3 | 4 | 2 | 2 | | 2 | 3 | 2 |
| Magelona filiformis | Р | 805 | 1 | | | | | | | | | | | |
| Magelona minuta | Р | 806 | | | | 2 | | | | | | | | 3 |
| Chaetopteridae | Р | 810 | | | | | | | | | | | | |
| Spiochaetopterus | Р | 820 | 14 | 5 | 2 | 10 | 6 | 9 | 11 | 9 | 6 | 6 | 13 | 4 |
| typicus | | | | | | | | | | | | | | |
| Cirratulidae | Р | 822 | | | | | | | | | | | | |
| Chaetozone setosa | Р | 834 | 9 | 14 | 9 | | 7 | 10 | 7 | | 3 | 11 | 20 | 4 |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|---------------------------|---|------|----|----|----|----|----|----|----|----|----|----|----|----|
| Chaetozone christiei | Р | | 2 | | | | | | | | | | | |
| Caulleriella killariensis | Р | 846 | | | | | | | | | 1 | | | |
| Tharyx sp. | Р | 847 | 14 | 17 | 10 | 7 | 6 | 4 | 9 | 14 | 6 | 9 | 26 | 9 |
| FLABELLIGERIDA | Р | 872 | | | | | | | | | | | | |
| Flabelligeridae | Р | 873 | | | | | | | | | | | | |
| Diplocirrus glaucus | Р | 878 | | 4 | 6 | 7 | 7 | 5 | 5 | 7 | 9 | 3 | 7 | 4 |
| CAPITELLIDA | Р | 902 | | | | | | | | | | | | |
| Capitellidae | Р | 903 | | | | | | | | | | | | |
| Mediomastus fragilis | Р | 919 | 3 | | 2 | | | 2 | 1 | | | | | 3 |
| Notomastus latericeus | Р | 921 | | 1 | | | | | 2 | | | 2 | 2 | |
| Maldanidae | Р | 938 | | | | | | | | | | | | |
| Euclymene lumbricoides | Р | 963 | | 3 | | | | | | | | | 3 | |
| OPHELIIDA | Р | 992 | | | | | | | | | | | | |
| Opheliidae | Р | 993 | | | | | | | | | | | | |
| Ophelina acuminata | Р | 1014 | | | | 1 | | | | | | | | |
| Scalibregmatidae | Р | 1020 | | | | | | | | | | | | |
| Scalibregma celticum | Р | 1026 | | | 1 | | | 2 | | | | | | |
| Scalibregma inflatum | Р | 1027 | 15 | | | 10 | 7 | | 7 | 23 | 8 | 6 | | |
| OWENIIDA | Р | 1089 | | | | | | | | | | | | |
| Oweniidae | Р | 1090 | | | | | | | | | | | | |
| Owenia fusiformis | Р | 1098 | 4 | 5 | | 4 | 3 | 2 | | 4 | | 7 | 5 | 3 |
| TEREBELLIDA | Р | 1099 | | | | | | | | | | | | |
| Pectinariidae | Р | 1100 | | | | | | | | | | | | |
| Pectinaria (Amphictene) | Р | 1102 | 2 | | | | | | | | | | | |
| auricoma | | | | | | | | | | | | | | |
| Ampharetidae | Р | 1118 | | | | | | | | | | | | |
| Melinna palmata | Р | 1124 | | | | | 1 | | 1 | | | | | |
| Ampharete sp. | Р | 1133 | | | | | | | | | | | | 8 |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|-----------------------|---|------|----|----|----|----|----|----|----|----|----|----|----|----|
| Ampharete lindstroemi | Р | 1139 | 7 | 8 | 11 | 4 | 3 | 9 | | 6 | | | | |
| Terebellidae | Р | 1179 | | | | | | | | | | | | |
| Lanice conchilega | Р | 1195 | | | | | | | | | | | 3 | |
| Polycirrus medusa | Р | 1242 | | 4 | | | | | | | 1 | | | |
| CRUSTACEA | R | 1 | | | | | | | | | | | | |
| AMPHIPODA | S | 97 | | | | | | | | | | | | |
| Oedicerotidae | S | 118 | | | | | | | | | | | | |
| Synchelidium | S | 138 | | | | | | | | | 1 | | 1 | |
| maculatum | | | | | | | | | | | | | | |
| Amphilochidae | S | 152 | | | | | | | | | | | | |
| Amphilochus | S | 159 | | | | | | | | 1 | | | | |
| neapolitanus | | | | | | | | | | | | | | |
| Leucothoidae | S | 175 | | | | | | | | | | | | |
| Leucothoe lilljeborgi | S | 178 | | | 1 | | | 1 | | | | | 2 | 1 |
| Phoxocephalidae | S | 252 | | | | | | | | | | | | |
| Harpinia sp. | S | 253 | | | | | | | 1 | | | | | |
| Harpinina serrata | S | 258 | 3 | 2 | | | | 4 | 4 | 7 | | | 7 | 4 |
| Lysianassidae | S | 271 | | | | | | | | | | | | |
| Orchomenella nana | S | 321 | | | | 1 | | | | | | | | |
| Synopioidea | S | 348 | | | | | | | | | | | | |
| Argissa hamatipes | S | 360 | 2 | 1 | | 1 | | | | 2 | | | | 1 |
| Ampeliscidae | S | 422 | | | | | | | | | | | | |
| Ampelisca brevicornis | S | 427 | 8 | 7 | 1 | | 9 | 2 | 6 | 9 | 6 | | 2 | 3 |
| Ampelisca spinipes | S | 438 | 7 | 3 | 1 | 3 | 5 | 1 | 3 | 4 | 2 | | 2 | 2 |
| Ampelisca tenuicornis | S | 440 | 1 | | 3 | 1 | | | | 1 | 1 | | | |
| Ampelisca typica | S | 442 | | | 1 | | | | | | | | | |
| Pontoporeiidae | S | 40 | | | | | | | | | | | | |
| Bathyporeia tenuipes | S | 459 | 1 | | | | | | | 1 | | | | |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|------------------------|---|------|----|----|----|----|----|----|----|----|----|----|----|----|
| Melitidae | S | 495 | | | | | | | | | | | | |
| Abludomelita obtusata | S | 498 | | | | | | 1 | | | | | | |
| Cheirocratus sp. | S | 503 | 1 | | | | | | | | | | 1 | |
| Melita sp. | S | 522 | | 1 | | | | | | | | | | |
| Melita dentata | S | 523 | | | 2 | | | | | | | | | |
| Isaeidae | S | 537 | | | | | | | | | | | | |
| Photis longicaudata | S | 552 | 2 | 1 | | 35 | 1 | | 2 | 5 | 2 | | 2 | |
| Caprellidae | S | 639 | | | | | | | | | | | | |
| Pariambus typicus | S | 651 | | | | | | | | 1 | | | 1 | |
| Phtisicidae | S | 655 | | | | | | | | | | | | |
| Phtisica marina | S | 657 | | | | | 1 | | | 2 | | | 3 | 9 |
| ISOPODA | S | 790 | | | | | | | | | | | | |
| Gnathiidae | S | 792 | | | | | | | | | | | | |
| Gnathia oxyuraea | S | 796 | | | | | 1 | | | | | | | |
| Paragnathia formica | S | 799 | | | | | | | | | | | 1 | |
| Arcturidae | S | 948 | | | | | | | | | | | | |
| Arcturella dilatata | S | 951 | | | | | | | | | | | 2 | 1 |
| TANAIDACEA | S | 1099 | | | | | | | | | | | | |
| Anarthruidae | S | 1115 | | | | | | | | | | | | |
| Tanaopsis graciloides | S | 1142 | 1 | | 3 | | | | | 3 | | | | |
| CUMACEA | S | 1183 | | | | | | | | | | | | |
| Bodotriidae | S | 1184 | | | | | | | | | | | | |
| Iphinoe serrata | S | 1201 | 1 | 1 | | 2 | | 2 | 2 | 3 | 5 | | 3 | 6 |
| Leuconiidae | S | 1204 | | | | | | | | | | | | |
| Eudorella truncatula | S | 1208 | 1 | | | 2 | | | 1 | 1 | 1 | | 3 | |
| Pseudocumatidae | S | 1231 | | | | | | | | | | | | |
| Pseudocuma longicornis | S | 1236 | | | | | | | | | | | 1 | 1 |
| Diastylidae | S | 1244 | | | | | | | | | | | | |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|-------------------------|---|------|----|----|----|----|----|----|----|----|----|----|----|----|
| Diastylis bradyi | S | 1248 | | | | | | | | 1 | 1 | | | |
| DECAPODA | S | 1276 | | | | | | | | | | | | |
| Processidae | S | 1361 | | | | | | | | | | | | |
| Processa sp. | S | 1362 | | 1 | | | | | | | 2 | | 2 | 1 |
| Processa nouveli | S | 1367 | 1 | | 2 | | | 1 | 3 | | | | 2 | |
| holthuisi | | | | | | | | | | | | | | |
| Crangonidae | S | 1380 | | | | | | | | | | | | |
| Philocheras trispinosus | S | 1390 | | | | | | | 1 | | | | | |
| Corystidae | S | 1550 | | | | | | | | | | | | |
| Corystes cassivelaunus | S | 1552 | | 1 | | | 1 | | | | | | | |
| MOLLUSCA | W | 1 | | | | | | | | | | | | |
| CHAETODERMATIDA | W | 3 | | | | | | | | | | | | |
| Chaetodermatidae | W | 7 | | | | | | | | | | | | |
| Chaetoderma nitidulum | W | 9 | | | | | | 1 | | | | | | |
| GASTROPODA | W | 88 | | | | | | | | | | | | |
| MESOGASTROPODA | W | 256 | | | | | | | | | | | | |
| Eulimidae | W | 599 | | | | | | | | | | | | |
| Eulima glabra | W | 604 | | | | | 2 | | 1 | | 2 | | | |
| NEOGASTROPODA | W | 670 | | | | | | | | | | | | |
| CEPHALASPIDEA | W | 1002 | | | | | | | | | | | | |
| Cylichnidae | W | 1024 | | | | | | | | | | | | |
| Cylichna cylindracea | W | 1028 | 2 | 4 | 1 | 2 | | 1 | 4 | 4 | | | 3 | 2 |
| Phylinidae | W | 1035 | | | | | | | | | | | | |
| Philine aperta | W | 1038 | 4 | | 3 | 3 | | | | | | | | 2 |
| PELECYPODA | W | 1560 | | | | | | | | | | | | |
| NUCULOIDA | W | 1561 | | | | | | | | | | | | |
| Nuculidae | W | 1563 | | | | | | | | | | | | |
| Nucula sp. | W | 1565 | | | | 3 | | | | | 3 | | | |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|----------------------|----|------|----|----|----|----|----|----|----|----|----|----|----|----|
| Nucula nitidosa | W | 1569 | | 1 | 1 | | | | | | | | | |
| VENEROIDA | W | 1815 | | | | | | | | | | | | |
| Lucinidae | W | 1817 | | | | | | | | | | | | |
| Myrtea spinifera | W | 1827 | | | | | | | | | 3 | | 2 | 2 |
| Thyasiridae | W | 1833 | | | | | | | | | | | | |
| Thyasira flexuosa | W | 1837 | 11 | 9 | 5 | 13 | | 6 | 7 | 9 | 12 | 9 | 9 | 3 |
| Montacutidae | W | 1888 | | | | | | | | | | | | |
| Tellimya ferruginosa | W | 1902 | | 2 | | | | 2 | 2 | 3 | 2 | | | |
| Mysella bidentata | W | 1906 | | | | | | | | 8 | | | | 2 |
| Mactridae | W | 1967 | | | | | | | | | | | | |
| Spisula subtruncata | W | 1978 | 2 | | | | | | | | | | | |
| Pharidae | W | 1995 | | | | | | | | | | | | |
| Phaxas pellucidus | W | 2006 | 10 | | 2 | 4 | | 2 | 5 | 7 | 9 | 7 | 7 | 8 |
| Psammobiidae | W | 2042 | | | | | | | | | | | | |
| Semelidae | W | 2057 | | | | | | | | | | | | |
| Abra alba | W | 2059 | | | | 1 | | | | | 1 | 2 | | |
| Abra nitida | W | 2061 | | | | | | | | | | | 1 | |
| Veneridae | W | 2086 | | | | | | | | | | | | |
| Chamelea striatula | W | | | | | 1 | | | | 2 | | 2 | 12 | |
| Timoclea ovata | W | 2104 | | | | | | 1 | | | | | | |
| Dosinia lupinus | W | 2128 | | | | | | | | | | | 1 | |
| MYOIDA | W | 2140 | | | | | | | | | | | | |
| Corbulidae | W | 2153 | | | | | | | | | | | | |
| Corbula gibba | W | 2157 | 2 | | 2 | 1 | | | 4 | 3 | | | | |
| PHORONIDA | ZA | 1 | | | | | | | | | | | | |
| Phoronidae | ZA | 2 | | | | | | | | | | | | |
| Phoronis sp. | ZA | 3 | 3 | 1 | | 2 | 2 | | | | 2 | | | |
| ECHINODERMATA | ZB | 1 | | | | | | | | | | | | |

| Station | | | 1A | 1B | 2A | 2B | 3A | 3B | 4A | 4B | 5A | 5B | 6A | 6B |
|------------------------|----|-----|----|----|----|----|----|----|----|----|----|----|----|----|
| OPHIUROIDEA | ZB | 105 | | | | | | | | | | | | |
| OPHIURIDA | ZB | 121 | | | | | | | | | | | | |
| Amphiuridae | ZB | 148 | | | | | | | | | | | | |
| Amphiura filiformis | ZB | 154 | 3 | 2 | 5 | 4 | 2 | | | 2 | 1 | 2 | | |
| Ophiuridae | ZB | 165 | | | | | | | | | | | | |
| Ophiura albida | ZB | 168 | | | 5 | 1 | 1 | | | | | | | |
| ECHINOIDEA | ZB | 181 | | | | | | | | | | | | |
| ECHINOIDA | ZB | 190 | | | | | | | | | | | | |
| Echinidae | ZB | 194 | | | | | | | | | | | | |
| Echinocyamus pusillus | ZB | 212 | | | | 1 | | | | | | | | |
| SPATANGOIDA | ZB | 213 | | | | | | | | | | | | |
| Loveniidae | ZB | 221 | | | | | | | | | | | | |
| Echinocardium | ZB | 223 | 2 | 3 | 3 | 3 | 2 | | 2 | 2 | 1 | 2 | 2 | |
| cordatum | | | | | | | | | | | | | | |
| HOLOTHURIOIDEA | ZB | 229 | | | | | | | | | | | | |
| DENDROCHIROTIDA | ZB | 249 | | | | | | | | | | | | |
| Cucumariidae | ZB | 266 | | | | | | | | | | | | |
| Leptopentacta elongata | ZB | 280 | 2 | | 6 | 3 | 1 | | | | 1 | 1 | 1 | |
| APODIDA | ZB | 289 | | | | | | | | | | | | |
| Synaptidae | ZB | 290 | | | | | | | | | | | | |
| Leptosynapta bergensis | ZB | 292 | 2 | 2 | 1 | 2 | 2 | | 2 | 1 | 1 | 2 | | |
| Labidoplax digitata | ZB | 300 | | | | | | | | | | | 1 | |

Appendix 2

Underwater Photographs

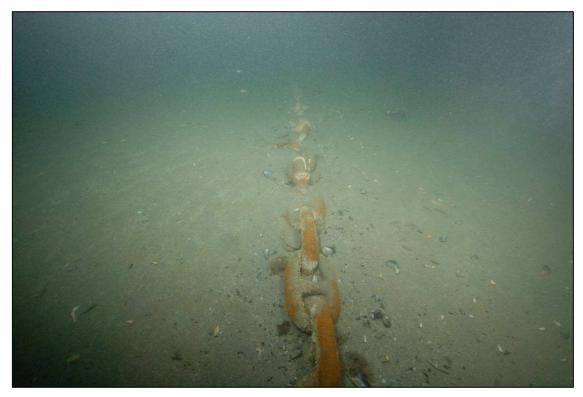


Figure 1: Anchor chain on mud at ocean energy device site

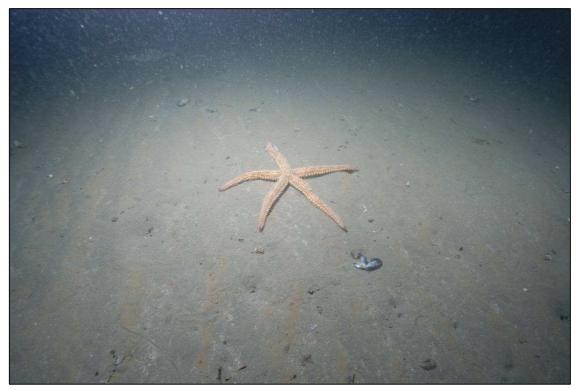


Figure 2: Marthasterias glacialis on muddy sand. Note presence of mussel shells



Figure 3: Goneplex rhomboides at ocean energy device site



Figure 4: Anchor chain with epifauna at ocean energy device site

Appendix 3

Sediment Profile Images

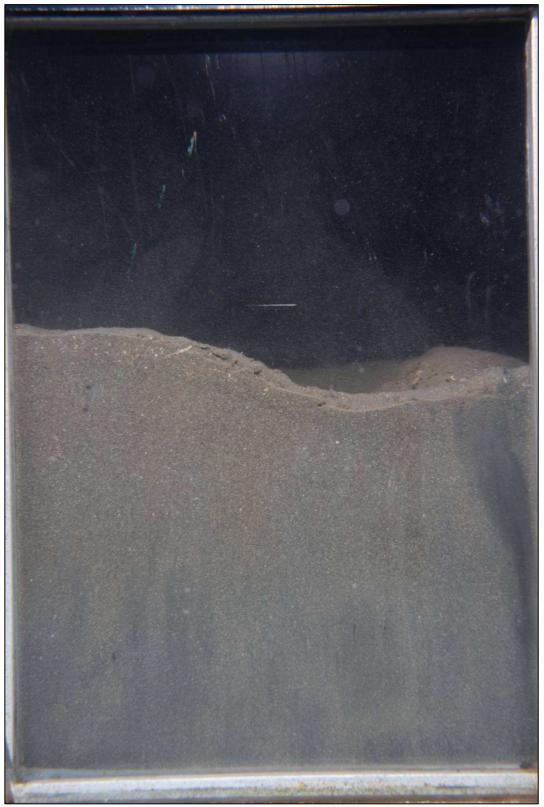


Figure 1: SPI image at ocean energy device site showing well oxygenated sediment with biologically generated surface boundary roughness.



Figure 2:SPI image showing a surface boundary which is not well defined